

## COMPLEXES BETWEEN POLYHYDROXY COMPOUNDS AND COPPER(II) IONS

E. J. BOURNE, F. SEARLE, AND H. WEIGEL

*Chemistry Department, Royal Holloway College (University of London), Englefield Green, Surrey (Great Britain)*

(Received May 28th, 1970; accepted for publication, June 29th, 1970)

### ABSTRACT

Several polyhydroxy compounds have been shown to form cationic complexes with copper(II) ions. Paper electrophoresis in copper(II) acetate and basic copper(II) acetate solutions, and chromatography on the  $\text{Cu}^{2+}$  form of a cation-exchange resin are useful methods for the resolution of mixtures and identification of polyhydroxy compounds.

### INTRODUCTION

The partially filled set of d-orbitals of the copper(II) ion confers, among other characteristic properties, the ability to form co-ordination complexes<sup>1</sup>. Pertinent examples of complexes with chelating ligands are those of glycosides of aldoses<sup>2</sup> and aminodeoxyaldoses<sup>3</sup> formed in cuprammonium solutions. In an effort to extend the range<sup>4</sup> of simple methods for the separation of carbohydrates and related compounds, we have investigated the paper electrophoresis in copper(II) acetate and basic copper(II) acetate solutions of a number of polyhydroxy compounds, and the chromatography of selected compounds on the  $\text{Cu}^{2+}$  form of a cation-exchange resin.

### RESULTS AND DISCUSSION

The compounds examined electrophoretically are listed in Table I. D-Glucitol (sorbitol) was used as a standard for the comparison of rates of migration, and 5-hydroxymethyl-2-furaldehyde as a non-migrating marker for correction of electro-osmosis. Hence, the migration rates are expressed as  $M_S$  values. The symbols  $\text{Cu}_{Ac}$  and  $\text{Cu}_{BAc}$  refer to copper(II) acetate and basic copper(II) acetate solutions, respectively. Migration was always towards the cathode.

It is impracticable to discuss in detail all the applications of this method of electrophoresis, since these will vary with the individual problems encountered. However, the results show that, in general, tetritols, pentitols, hexitols, and reduced disaccharides of D-glucose (except laminaribitol) form cationic complexes with the copper(II) ion, but, under the conditions described here, reducing sugars (except D-ribose, D-xylose, and D-gulose) do not. It is further interesting to note that each of

TABLE I

ELECTROPHORETIC MOBILITIES OF ALDITOLS IN COPPER(II) ACETATE AND BASIC COPPER(II) ACETATE SOLUTIONS

| <i>Compound</i> | $M_S$ (CuAc) | $M_S$ (CuBAc) | <i>Non-migrating compounds:<br/><math>M_S</math> (CuAc) and <math>M_S</math> (CuBAc) &lt; 0.1</i> |
|-----------------|--------------|---------------|---|
| Erythritol      | 0.24         | <0.1          | Glycerol  |
| L-Threitol      | 0.7–0.9      | 0.20          | L-Arabinose   |
| L-Arabinitol    | 0.5–1.0      | 0.87          | D-Lyxose  |
| Ribitol         | 0.20         | 0.17          | D-Galactose   |
| Xylitol         | 1.35         | 1.20          | D-Glucose   |
| Allitol         | 0.25         | 0.36          |   |
| D-Altritol      | 0.5–1.0      | 0.89          | D-Mannose   |
| Galactitol      | 1.03         | 1.00          | D-Fructose  |
| D-Glucitol      | 1.00         | 1.00          | L-Sorbose   |
| L-Iditol        | 0.6–1.3      | 1.15          | Sucrose   |
| D-Mannitol      | 1.00         | 0.88          | Kojibiose   |
| Kojibitol       | 0.53         | 0.39          | Sophorose   |
| Sophoritol      | 0.30         | 0.29          | Nigerose  |
| Nigeritol       | 0.34         | 0.16          | Laminaribiose   |
| Maltitol        | 0.72         | 0.54          | Maltose   |
| Cellobiitol     | 0.27         | 0.18          | Cellobiose  |
| Isomaltitol     | 0.88         | 0.72          | Isomaltose  |
| Gentiobitol     | 0.92         | 0.88          | Gentiobiose   |
| D-Ribose        | 0.15         | <0.1          | Laminaributol   |
| D-Xylose        | 0–0.3        | <0.1          |   |
| D-Gulose        | 0.37         | <0.1          |   |

the four pairs of  $\alpha$ - and  $\beta$ -isomers of reduced disaccharides of D-glucose can be resolved by this method.

It is tedious to apply electrophoretic techniques to preparative work, unless a special apparatus is used. However, that complexing of polyhydroxy compounds with copper(II) ions can easily be used when larger quantities of materials are involved is illustrated by the successful resolution of mixtures on Amberlite IR-120 (Cu<sup>2+</sup>) resin (Table II). The results indicate that chromatography on this resin resolves mixtures which can also be resolved by paper electrophoresis in copper(II) acetate or basic copper(II) acetate solutions.

#### EXPERIMENTAL

*Paper electrophoresis.* — Electrophoresis was carried out on sheets (10 cm wide) of Whatman No. 3 filter paper. The electrolytes consisted of freshly prepared 5% aqueous copper(II) acetate monohydrate, Cu(CH<sub>3</sub>CO<sub>2</sub>)<sub>2</sub>·H<sub>2</sub>O (pH 5.1), or 5% aqueous basic copper(II) acetate, Cu(CH<sub>3</sub>CO<sub>2</sub>)<sub>2</sub>·CuO·6H<sub>2</sub>O (pH 5.1–5.3). Electrophoretograms were prepared by applying a voltage of *ca.* 60 volts/cm for *ca.* 2 h. Compounds were detected by treating the dried paper with a saturated solution of potassium permanganate in acetone. Under the conditions used, D-glucitol had mobilities of  $2.65 \times 10^{-5}$  and  $2.21 \times 10^{-5}$  cm<sup>2</sup> volt<sup>-1</sup>sec<sup>-1</sup> in aqueous copper(II) acetate and aqueous basic copper(II) acetate, respectively.

TABLE II

CHROMATOGRAPHY OF ALDITOLS ON AMBERLITE IR-120(Cu<sup>2+</sup>) RESIN

| <i>Components of mixture</i> | <i>Weight (mg)</i> | <i>Size of column (cm)</i> | <i>Eluent<sup>a</sup> (ml)</i>                | <i>Effluent containing pure sample (ml)</i> | <i>Extent of separation</i> | <i>Recovery (mg)</i> |
|------------------------------|--------------------|----------------------------|---|---|-----------------------------|----------------------|
| D-Glucose                    | 200                | 4 × 33                     | 730(10) <sup>c</sup> , 1080(10) <sup>g</sup>  | 31–210                                      | Complete                    |                      |
| D-Glucitol                   | 200                |                            |   | 1611–1810                                   |                             |                      |
| D-Mannose                    | 100                | 2.5 × 36                   | 3000(50) <sup>b</sup> , 500(500) <sup>g</sup> | 1–1500                                      | Complete                    |                      |
| D-Mannitol                   | 100                |                            |   | 3000–3500                                   |                             |                      |
| Cellobiitol                  | 100                | 2.5 × 36                   | 1750(25) <sup>d</sup>                         | 1–500                                       | Partial                     |                      |
| Maltitol                     | 100                |                            |   | 1000–1750                                   |                             |                      |
| Maltitol                     | 100                | 4 × 33                     | 1850(10) <sup>e</sup> , 450(10) <sup>f</sup>  | 51–1850                                     | Complete                    | 80                   |
| Isomaltitol                  | 100                |                            | 2000(100) <sup>g</sup>                        | 2051–4300                                   |                             | 95                   |
| Ribitol                      | 200                |                            |   | 1–100                                       | Partial                     | 132                  |
| D-Arabinitol                 | 200                | 2.5 × 36                   | 1625(25) <sup>e</sup>                         | 251–700                                     | Partial                     | 107                  |
| Xylitol                      | 200                |                            |   | 976–1625                                    | Partial                     | 122                  |

<sup>a</sup>Figures in parenthesis are the volumes (ml) of individual fractions collected; <sup>b</sup>water; <sup>c</sup>0.2% CuAc<sub>2</sub> · H<sub>2</sub>O; <sup>d</sup>0.4% CuAc<sub>2</sub> · H<sub>2</sub>O; <sup>e</sup>1% CuAc<sub>2</sub> · H<sub>2</sub>O; <sup>f</sup>1.25% CuAc<sub>2</sub> · H<sub>2</sub>O; <sup>g</sup>2.5% CuAc<sub>2</sub> · H<sub>2</sub>O.

*Chromatography.* — Columns of Amberlite IR-120 (H<sup>+</sup>) resin were treated with 2.5% aqueous copper(II) acetate monohydrate (2–4 l). The mixtures of polyhydroxy compounds dissolved in 2.5% aqueous copper(II) acetate monohydrate (2 ml) were run into the columns and the whole allowed to stand overnight. The effluents were analysed by paper electrophoresis in copper(II) acetate solution (see above). Fractions from which the solute was recovered were treated with gaseous hydrogen sulphide, filtered, and evaporated under reduced pressure. Other details are shown in Table II.

## ACKNOWLEDGMENTS

The authors thank the Science Research Council for financial assistance.

## REFERENCES

- 1 L. E. ORGEL, *An Introduction to Transition Metal Chemistry*, Methuen, London, 1960, p. 11.
- 2 R. E. REEVES, *Advan. Carbohydr. Chem.*, 6 (1951) 107.
- 3 C. B. BARLOW AND R. D. GUTHRIE, *J. Chem. Soc. (C)*, (1967) 1194.
- 4 H. WEIGEL, *Advan. Carbohydr. Chem.*, 18 (1963) 61.